Gil Nelson: https://www.idigbio.org/wiki/index.php/Fluid_Preserved_Arthropds

Sandra Brantley: Not my best slides. ;-) I think those are eggs in the body.

Mariel Campbell: These are wonderful - the specimen isn't preserved well, but the image is fantastic.

Elspeth Haston: What are the relative image file sizes in total? Do you keep all the stacks or only the final composite version?

Mariel Campbell: What is the maximum magnfication on the Hamamatsu?

Sandra Brantley: I am leaning toward archiving the final composite image and not the stacks but haven't made a decision yet.

Elspeth Haston: What about slides with multiple specimens, eg diatoms?

Robin Delapena: Could this be used to image a shell .5mm in size

Gisela Canales: yes

Tiffany Adrain: What's the largest slide size that can be scanned? I'm thinking about fossil coal ball peels (not entirely flat sections that could benefit from image stacking, maybe)

elisa: i missed how long it takes to scan 200 slides, has anyone cought it?

Mark Metz: Can the scannerr output formats other than .jpg for the stacks?

Scott Blakely: This is Scott Blakely with Hamamatsu: Great question. The NanoZoomer HT scans 15mmx15mm at 20X in 1 minute. 2.5 minutes at 40X. This is brightfield only, and this would be per Z section. The NanoZoomer XR is more than twice as fast as the HT in brightfield.

Mark Metz: If the stack files are proprietary, don't bother archiving them unless you can convert to an open standard.

Sandra Brantley: Right - that's why I'm thinking of just keeping the composite tiff.

Scott Blakely: Mark: The default native image is called an NDPI. We can then export this using NDP.Toolkit to TIFF or Jpeg. There is a 4Gbyte maximum file size built into the TIFF specification.

Scott Blakely: Tiffany:The NanoZoomer HT and XR can scan standard 1x3 inch slides. The NanoZoomer RS can also scan 2x3 inch slides.

Elspeth Haston: Elspeth Haston: e.haston@rbge.org.uk

Gil Nelson: rdm@meyerinst.com

Mark Metz: Can you control Z-depth per slide or is it overall for a 200 slide tray?

Robin Delapena: yes

Tiffany Adrain: Thanks, Scott answered my question!

Mark Metz: What kind of control is available for the light source?

Mark Metz: Can you change intensity, color, angle of incidence?

Felipe Soto: What about unstained specimens? springtails are not stained and we use phase contrast to be able to see the setae

Scott Blakely: Mark: I am struggling with getting my michrphone to work. Can you clarify this question? What would you like to do?

Mark Metz: The examples Rob showed had a lot of light artifact, need condensed/focused light.

Sandra Brantley: Felipe: phase contrast also used for mites, but the setal fields came out well on the mite images.

Felipe Soto: mites have a tinge and tend to look better than springtails, even without contrast

Robin Delapena: can you elaborate on the "blending" of the three rendering options to get better results

Robin Delapena: yes please! rdelapena@fieldmuseum.org

Gil Nelson: https://www.idigbio.org/wiki/index.php/Fluid_Preserved_Arthropds

Gisela Canales: I am taking notes for you

Jim Woolley: Thanks for this Rob, by the way, check out ZereneStacker for z-stacking, we have totally switched to that software in my lab

Robin Delapena: Photoshop blending techniques - rdelapena@fieldmuseum.org (Robin Delapena)

Mark Metz: Thanks, Rob and Scott.

Mariel Campbell: Thank you so much!

Robin Delapena: Thanks guys!!

Kal Ivanov: Thanks

Sandra Brantley: Thank you